

Allele frequencies of 163A/C polymorphism of the *CYP1A2* gene in the selected Ukrainian population

¹Filipitsova O.V., ²Kobets M.N., Kobets Yu.N., ¹Naboka O.I., ¹Timoshyna I.A., ²Galiy L.V.

National University of Pharmacy, Biology Department, Ukraine, Kharkov

National University of Pharmacy, Pharmaceutical Marketing and Management Department, Ukraine, Kharkov

Abstract

Background: *CYP1A2* is a very important gene for a potential genotyping. The objective of this work was to study the population frequencies of the corresponding 163A/C single nucleotide polymorphism of the *CYP1A2* gene in the sampling of the Ukrainian population.

Methods: Total sampling of genetic material (buccal epithelium) was collected in 102 subjects (48 males, 54 females) **not related** to each other. Genotype test of the participants of the study for *CYP1A2* polymorphism (rs762551) was performed using the polymerase chain reaction. In accordance with the findings of genotyping the allele frequencies (p and q) were calculated.

Results: The following distribution of genotypes in a sampling of the Ukrainian population under the 163A/C polymorphism of the *CYP1A2* gene: AA - 36%, AC - 49% and CC - 15% in the subjects was revealed. The population allele frequencies of the 163A/C polymorphism of the *CYP1A2* gene totaled $p_A = 0.6$ and $q_C = 0.4$. The population structure of the individuals does not deviate from the Hardy-Weinberg equilibrium, as may be seen from the lack of difference between the theoretically expected and the observed frequencies of the three genotypes.

Conclusion: The genetic polymorphism found in the Ukrainian population is the basis for the recommendation of the genetic testing of 163A/C polymorphism of the *CYP1A2* gene in the prescribing **drugs - substrates** of this gene.

Keywords: *CYP1A2* (163A/C polymorphism), population distribution, Ukraine.

Introduction

The *CYP1A2* gene is located in chromosome 15 **and composed** of 7 exons and 6 introns (1). *CYP1A2* is one of the major liver enzymes, which is involved in the metabolism of the variety of drugs, **widespread** suspected carcinogens and endogenous compounds. As the cytochrome is expressed in the liver, it can metabolize the compounds without additional induction. Among the drugs, which undergo a substantial (over 30%) metabolic transformation under its influence, are, in particular, caffeine, theophylline, tacrine, melatonin, verapamil, clozapine, olanzapine, aminopyrine, propranolol, etc. The drugs, that to a greater or lesser (30 to 10%) **extent are** exposed to the action of *CYP1A2*, include acetaminophen, lidocaine, imipramine, antipyrin, etc. Examples of the suspected carcinogens activated by ***CYP1A2*** are benzopyrene and aflatoxin B1, and the examples of the endogenous compounds are steroids. *CYP1A2* has a number of inducers and suppressors, which can temporarily alter its genetically determined activity. **Some of allele forms of *CYP1A2*** are particularly sensitive to the effect of the respective inducers (2).

So far, there are more than 30 variants of *CYP1A2* allelic forms. In particular, during the study of DNA samples taken from Japanese individuals, the most common single nucleotide polymorphism was 163A/C (rs762551), which is defined as an allele *CYP1A2*1F* (3). In one study it was offered to define the allele as the C allele of the wild-type, and the A allele — as a mutant variant (4). It is also known, that from the several variants of the gene *CYP1A2* allele *CYP1A2*1F* is marked by a particularly high **inducibility** (5). *CYP1A2*1F* is responsible for the shape of the enzyme with poor metabolic activity (2). For example, earlier studies **have shown**, that smoking increased the activity of the enzyme *CYP1A2*. On this inducing influence an increase in the metabolic ratio of caffeine was clearly indicated in smokers. In the males – inveterate smokers, the enzyme activity was also correlated with the metabolism of acetaminophen, which is partly manifested in the **glucuronidation** of the compound (6).

The study is known, that examined the effect of the *CYP1A2* gene polymorphism on early aging due to industrial hazards. The results of **one** study pointed at a higher level of DNA damage in the cells and, consequently, their premature deterioration and aging, that had a mutant form of *CYP1A2* and were exposed to harmful compounds (7).

As the rs762551 genetic polymorphism under the gene, encoding CYP1A2, affects the inducibility of the expression of *CYP1A2* and may be associated with different types of cancer. One of the meta-analyses, that included 19 studies with a "case-control" design, illustrated the probable association of *CC* genotype with an increased risk of malignant neoplasms in the Caucasians. In the mixed populations and in the Asians such relationship was not found (8). In a more recent meta-analysis, that summarized 71 studies, the relationship between the variants of the ***CYP1A2* gene** and predisposition to cancer was not **found** (9).

The 163A/C polymorphism of the *CYP1A2* gene is associated with many common human diseases. In particular, this polymorphism can play a greater role in the predisposition to chronic diseases of the respiratory tract considerably caused by toxic substances, for example, the components of tobacco smoke, that get into the lungs. It was shown that the *C* allele frequency in patients with chronic obstructive pulmonary disease was higher in comparison with the control group. It was concluded that the polymorphism of the *CYP1A2* gene may be associated with both **treatment efficacy** with the bronchodilator theophylline, and the predisposition to chronic obstructive pulmonary disease (10). A similar study was conducted earlier in Bashkortostan, but the relationship between the polymorphism of the *CYP1A2* gene and predisposition to chronic obstructive pulmonary disease was not detected (11).

At the same time the study of treatment efficacy with **betaxolol**, a synthetic antihypertensive drug from the group of β_1 -blockers, has shown that the therapeutic effect, to a greater extent, had been achieved for the poor metabolizers – subjects with *CC* genotype of 163A/C variant of the *CYP1A2* gene. The authors of the study attributed this to the fact, that the metabolism of the drug had been reduced, and its therapeutic effect on the patients was longer. The observed effect **was shown in the** Russian population (12).

It was also shown, that in poor *CC* **metabolizers** clearance of **teriflunomide**, which is the active metabolite of the anti-rheumatic drug **leflunomide**, was slightly lower, than in the extensive (*AA*) and intermediate (*AC*) metabolizers. However, the noted phenomenon was only a trend (13).

CYP1A2 is a very important gene for a potential genotyping due to all described above **and** it was not yet been studied by any polymorphism in the population of Ukraine. In our previous research we have demonstrated that understanding of **pharmacogenetics** is different in males and females, but it is highly desirable it be increasing (14). Even in this situation some part of the Ukrainians is ready to pay for genetic tests which are part of personalized medicine (15). The objective of this work was to study the population frequencies of the corresponding 163A/C single nucleotide polymorphism of the *CYP1A2* gene in the sampling of the Ukrainian population.

Materials and Methods

To study the population distribution of the 163A/C polymorphism of the *CYP1A2* gene a **sampling consisting of the Ukrainian individuals was formed**. It should be noted that ethnicities of **people living** in the area of the modern Ukraine are predominantly Ukrainians and Russians with different proportions in the different locations of the country (16-18). Total sampling of genetic material (buccal epithelium) was collected in 102 subjects (48 males, 54 females), who were not related to each other. **Participants were predominantly healthy students of National University of Pharmacy, aged 17 to 25. Material was collected in accordance with ethical standards of work under Helsinki Declaration (World Medical Association Declaration of Helsinki, Ethical Principles for Medical Research Involving Human Subjects).** Genotype test of the participants of the study for *CYP1A2* polymorphism (rs762551) was performed using the polymerase chain reaction (19).

The DNA was isolated from the test samples of buccal epithelium using the ion exchange resin Chelex-100 (20). Determination of the allelic states of the *CYP1A2*1F* gene under the single nucleotide replacement of *CYP1A2* (rs762551) was carried out according to the standard methods (4). The amplification was performed on the "Tertsik" thermocycler (amplifier) (DNA-Technology, Russia).

For the fragment amplification of the *CYP1A2* gene, which contained the polymorphic site (163A/C) Oligonucleotide primers were used: the forward CCCAGAAGTGGAACTGAGA and the reverse GGGTTGAGATGGAGACATTC (4). The restriction of the amplification products was performed using the *ApaI* endonuclease (MBI Ferments, Lithuania). The restriction of amplification products was analyzed with electrophoresis in 2% agarose gel. As the marker of molecular weight the DNA pUC19 was used, hydrolyzed with the *MspI* endonuclease (MBI Ferments, Lithuania). The visualization of the amplification and restriction products were carried out by the staining with ethidium bromide and photographing at the transilluminator under the UV light. The restriction fragment of 243 bp size corresponded to A allele under the variant 163A/C of the *CYP1A2* gene, and two restriction fragments of 119 and 124 bp size corresponded to the C allele. The presence of all three bands at the electrophoregram indicated at the heterozygous AC genotype (4).

In accordance with the findings of genotyping the allele frequencies (p and q) were calculated:

$$p_A = \frac{2AA + AC}{2N} \quad \text{and} \quad q_C = \frac{2CC + AC}{2N},$$

where N – is a number of subjects.

The testing of the genotypes distribution on Hardy-Weinberg equilibrium using the criterion χ^2 was performed. The statistical hypothesis testing at $p \leq 0.05$ level of significance was conducted.

Results

Figure 1 shows the findings of electrophoretic fractionation in 2% agarose gel of human DNA amplified in PCR and hydrolyzed by endonuclease *ApaI*.

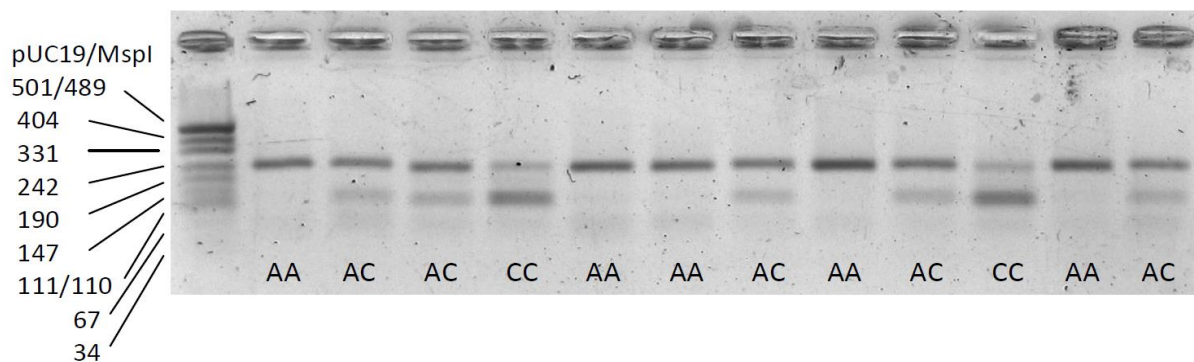


Figure 1. The electrophoretic fractionation in 2% agarose gel of the human DNA amplified PCR and hydrolyzed by endonuclease *ApaI*: first marker-band- *pUC19/MspI*, 2-13 marker-bands - DNA of the subjects.

Genotype test of the subjects for the 163A/C polymorphism of the *CYP1A2* gene found, that in the studied sampling the fewest were poor (CC, 15 of 102), and the most were intermediate (AC, 50 of 102) metabolizers. In general, in the population sampling the percentage distribution of the genotypes was as follows: AA — in 36%, AC — in 49% and CC - in 15% subjects (Table 1).

Table 1

The distribution of genotypes of 163A/C polymorphism of *CYP1A2* gene

	Males, n	Females, n	Total, N (%)
AA	19	18	37 (36)

<i>AC</i>	21	29	50 (49)
<i>CC</i>	8	7	15 (15%)

Statistics: $\chi^2 = 1.024$, $df = 2$, $p > 0.05$.

Note. χ^2 – Pearson criterion, df – degree of freedom, p – significance level.

Separately for males and females the allelic frequencies *A* and *C* were calculated. The weighted average frequencies of corresponding alleles which totaled: $p_A = 0.6$ and $q_C = 0.4$, respectively were also calculated (Table 2).

Table 2

The allelic frequency of *A* and *C* of *CYP1A2* gene (163A/C polymorphism)

	Alleles	
	<i>A</i>	<i>C</i>
Males	0.61	0.39
Females	0.6	0.4
Total	0.6	0.4

According to the allele frequencies, on the basis of Hardy-Weinberg proportions were calculated the frequencies of the respective genotypes (Table 3).

Table 3

The genotype frequencies under the 163A/C polymorphism of the *CYP1A2* gene

	Genotypes		
	<i>AA</i>	<i>AC</i>	<i>CC</i>
Males	0.37	0.48	0.15
Females	0.36	0.48	0.16
Total	0.36	0.48	0.16

The theoretically expected genotype frequencies calculated from the Hardy-Weinberg equilibrium was not significantly different from the observed ones (Table 4). This makes it possible to conclude about equilibrium of the 163A/C polymorphism of the *CYP1A2* gene in a sampling of the Ukrainian population.

Table 4

The theoretically expected population genotype frequencies of the polymorphism 163A/C of the gene *CYP1A2*

	Theoretically expected genotype frequencies	Observed genotype frequencies
<i>AA</i>	37	37
<i>AC</i>	49	50
<i>CC</i>	16	15

Statistics: $\chi^2 = 0.042$, $df = 2$, $p > 0.05$.

Note. The designations are the same as for Table 1.

Discussion

The frequencies of these alleles were studied in several populations. The studies have shown that the world's population is variable.

The study the 163A/C polymorphism distribution of the *CYP1A2* gene in 159 healthy Japanese individuals found, that *CC* homozygotes occurred at a frequency of 16.4%, homozygotes *AA* – 39% and heterozygotes *AC* – 44.6% (4). Among the 71 patients with rheumatoid arthritis at the age from 27 to 82, residents of Central Europe and being treated in the clinic of Slovenia *CC* genotype was present in 5.6%, *AC* in 56.3% and *AA* in 38% of the subjects, respectively. More detailed information about the origin of the subjects was not available (13). Among the healthy Turkish population ($n = 101$) and the patients with chronic

obstructive pulmonary disease, the subjects with *CC* genotype occurred at a frequency of 6.9% and 11%, *AC* – 39.6% and 58%, and *AA* – 53.5% and 31%, respectively. The respective allelic frequencies in healthy people were $p_A = 0.73$ and $q_C = 0.27$, and in patients – $p_A = 0.6$ and $q_C = 0.4$ (10).

The frequencies of different genotypes of the 163A/C polymorphism of the *CYP1A2* gene in other Slavic populations can be assessed by the study conducted by Russian scientists. In particular, there is available information on the **genotypes distribution** in a sampling of the patients with cardiovascular diseases – hypertension and atrial fibrillation, with a total of 81 people. Thus, in the various subgroups separated depending on objectives of the study, the frequency of poor *CC* metabolizers was 8% and 27%. The extensive (*AA*) and intermediate (*AC*) metabolizers were 68%/44% and 24%/29%, respectively. The allele frequencies in different subgroups varied from 0.59 to 0.8, and *C* allele – from 0.2 to 0.41, respectively (12).

Thus, the data provided allowed to make a **suggestion that** the studied gene frequencies and, respectively, the frequencies of different genotypes, point at the presence of intra-population differences in the 163A/C polymorphism of the *CYP1A2* gene (Figure 2), **yet the small sample size should be defined as limitation of our study.**

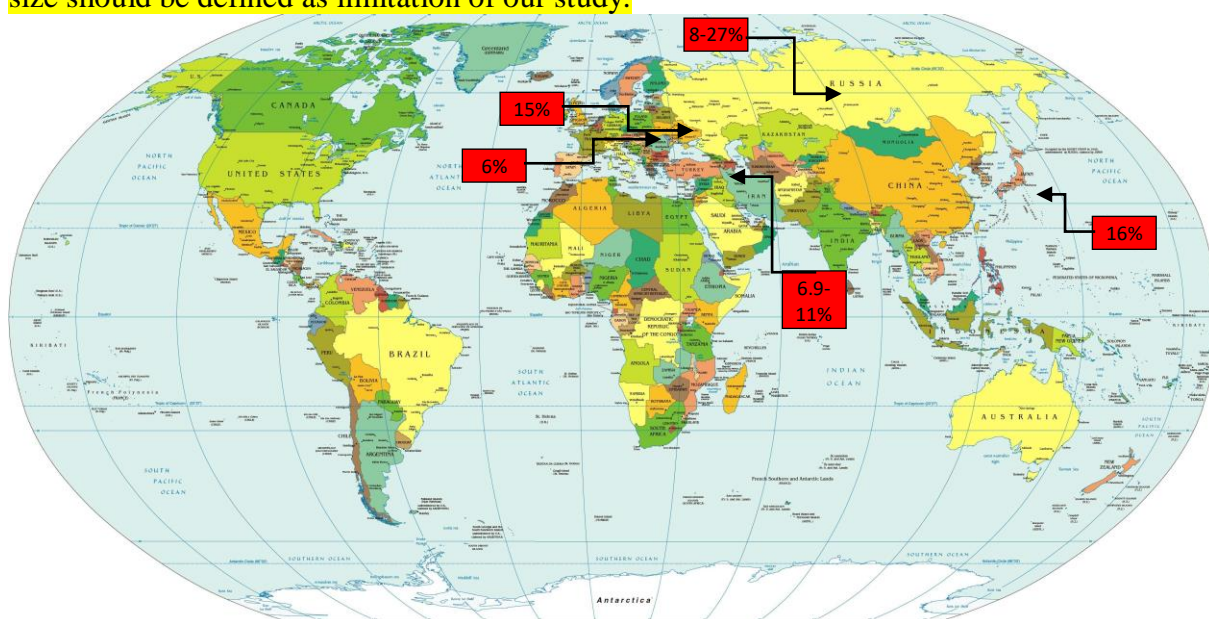


Figure 2. The frequency of poor metabolizers and *CC* homozygotes under the 163A/C polymorphism of the *CYP1A2* gene (according to the references (see links in the text) and according to the original survey).

Conclusions

1. The following distribution of genotypes in a sampling of the Ukrainian population under the 163A/C polymorphism of the *CYP1A2* gene: *AA* – 36%, *AC* – 49% and *CC* – 15% in the subjects was revealed.
2. The population allele frequencies of the 163A/C polymorphism of the *CYP1A2* gene totaled $p_A = 0.6$ and $q_C = 0.4$.
3. The population structure of the individuals does not deviate from the Hardy-Weinberg equilibrium, as may be observed from the lack of difference between the theoretically expected and the frequencies of the three genotypes.
4. The genetic polymorphism found in the Ukrainian population is the basis for the recommendation of the genetic testing of 163A/C polymorphism of the *CYP1A2* gene in the prescribing **drugs that** are the substrates of this gene.

Acknowledgments

The research was partially carried out with the support of the grant of the Ministry of Education and Science of Ukraine "Biological Challenges and Threats due to Migrations and Invasions: a Population-Genetic Approach" (2017).

References

1. [Ikeya K](#), [Jaiswal AK](#), [Owens RA](#), [Jones JE](#), [Nebert DW](#), [Kimura S](#). Human CYP1A2: sequence, gene structure, comparison with the mouse and rat orthologous gene, and differences in liver 1A2 mRNA expression. [Molecular Endocrinology](#), 1989; 3:1399-408.
2. [Zhou SF](#), [Yang LP](#), [Zhou ZW](#), [Liu YH](#), [Chan E](#). Insights into the substrate specificity, inhibitors, regulation, and polymorphisms and the clinical impact of human cytochrome P450 1A2. [The AAPS Journal](#), 2009; 11:481-94.
3. [Soyama A](#), [Saito Y](#), [Hanioka N](#), [Maekawa K](#), [Komamura K](#), [Kamakura S](#), [Kitakaze M](#), [Tomoike H](#), [Ueno K](#), [Goto Y](#), [Kimura H](#), [Kato M](#), [Sugai K](#), [Saitoh O](#), [Kawai M](#), [Ohnuma T](#), [Ohtsuki T](#), [Suzuki C](#), [Minami N](#), [Kamatani N](#), [Ozawa S](#), [Sawada J](#). Single nucleotide polymorphisms and haplotypes of CYP1A2 in a Japanese population. [Drug Metabolism and Pharmacokinetics](#), 2005; 20:24-33.
4. [Chida M](#), [Yokoi T](#), [Fukui T](#), [Kinoshita M](#), [Yokota J](#), [Kamataki T](#). Detection of three genetic polymorphisms in the 5'-flanking region and intron 1 of human CYP1A2 in the Japanese population. [Japanese Journal of Cancer Research](#), 1999; 90:899-902.
5. [Ghotbi R](#), [Christensen M](#), [Roh HK](#), [Ingelman-Sundberg M](#), [Aklillu E](#), [Bertilsson L](#). Comparisons of CYP1A2 genetic polymorphisms, enzyme activity and the genotype-phenotype relationship in Swedes and Koreans. [The European Journal of Clinical Pharmacology](#), 2007; 63:537-46.
6. [Bock KW](#), [Schrenk D](#), [Forster A](#), [Griese EU](#), [Mörike K](#), [Brockmeier D](#), [Eichelbaum M](#). The influence of environmental and genetic factors on CYP2D6, CYP1A2 and UDP-glucuronosyltransferases in man using sparteine, caffeine, and paracetamol as probes. [Pharmacogenetics](#), 1994; 4:209-18.
7. [Eshkoor S](#), [Ismail P](#), [Rahman S](#), [Moin S](#), [Adon M](#). Role of the Gene Polymorphism on Early Ageing from Occupational Exposure. [The Balkan Journal of Medical Genetics](#), 2013; 16:45-52.
8. [Wang H](#), [Zhang Z](#), [Han S](#), [Lu Y](#), [Feng F](#), [Yuan J](#). CYP1A2 rs762551 polymorphism contributes to cancer susceptibility: a meta-analysis from 19 case-control studies. [BMC Cancer](#), 2012; 12:528.
9. [Vukovic V.](#), [Ianuale C.](#), [Leoncini E.](#), [Pastorino R.](#), [Gualano MR](#), [Amore R](#), [Boccia S](#). Lack of association between polymorphisms in the CYP1A2 gene and risk of cancer: evidence from meta-analyses. [BMC Cancer](#), 2016; 16:83.
10. [Uslu A](#), [Ogus C](#), [Ozdemir T](#), [Bilgen T](#), [Tosun O](#), [Keser I](#). The effect of CYP1A2 gene polymorphisms on Theophylline metabolism and chronic obstructive pulmonary disease in Turkish patients. [BMB Reports](#), 2010; 43:530-4.
11. [Korytina GF](#), [Akhmadishina LZ](#), [Kochetova OV](#), [Zagidullin ShZ](#), [Viktorova TV](#). Association of cytochrome P450 genes polymorphisms (CYP1A1 and CYP1A2) with the development of chronic obstructive pulmonary disease in Bashkortostan. [Molecular Biology \(Mosk\)](#). 2008; 42:32-41.
12. Mynushkyna LO, Horshkova ES, Mankhaeva BB, Savel'eva EH, Kochkyna MS, Brovkyn AN, Nykytyn AH, Zateyshchikova AA, Nosykov VV, Zateyshchikov DA. Henetycheskye aspekty indyvydual'noy chuvstvyytel'nosti k betaksololu u bol'nykh hypertonycheskoy bolezn'yu y mertsatel'noy arytymyey. [Kremlevskaya medytsyna. Klynycheskyy vestnyk](#). 2014; 2:20-5 (in Russian).
13. [Bohanec Grabar P](#), [Grabnar I](#), [Rozman B](#), [Logar D](#), [Tomsic M](#), [Suput D](#), [Trdan T](#), [Peterlin Masic L](#), [Mrhar A](#), [Dolzan V](#). Investigation of the influence of CYP1A2

and CYP2C19 genetic polymorphism on 2-Cyano-3-hydroxy-N-[4-(trifluoromethyl)phenyl]-2-butenamide (A77 1726) pharmacokinetics in leflunomide-treated patients with rheumatoid arthritis. [Drug Metabolism and Disposition](#), 2009; 37:2061-8.

14. Filiptsova OV, Kobets MN, Kobets YuN. Some aspects of genetics and pharmacogenetics understanding by pharmacy students in Ukraine. *The Egyptian Journal of Medical Human Genetics*, 2015. 16(1):61-6.
15. Filiptsova O, Naboka O, Kobets M, Kobets Yu. Pharmacogenetic Tests in Ukraine: Economic Aspect. *Gazi Medical Journal*, 2017; 28(2):79-84.
16. Atramentova LA, Filiptsova OV. Genetic Demographic Processes in Ukrainian Urban Populations in 1990s: The Marriage Structure of the Kharkov Population. *Russian Journal of Genetics*, 1998; 34(8): 941-46.
17. [Atramentova LA](#), [Filiptsova OV](#). Genetic Demographic Processes in Ukrainian Urban Populations in the 1990s: The Marriage Structure of the Poltava Population. *Russian Journal of Genetics*, 1999; 35(12):1464-70.
18. Atramentova LA, Filiptsova OV, Mukhin VN, Osipenko SYu. Genetic Demography of Ukrainian Urban Populations in the 1990s: Ethnic Geographic Characteristics of Migration in the Donetsk Population. *Genetika*, 2002; 38(10):1402-8.
19. [Garibyan L](#), [Avashia N](#). Research Techniques Made Simple: Polymerase Chain Reaction (PCR). [Journal of Investigative Dermatology](#), 2013; 133(3):e6.
20. Walsh PS, Metzger DA, Higuchi R. Chelex 100 as a medium for extraction of DNA for PCR-based typing from forensic material. *BioTechniques*, 1991; 10:506-13.